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Cytotoxic 8,9-Secokaurane Diterpenes from a New Zealand Liverwort, Lepidolaena taylorii

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Abstract: The main cytotoxic compound from the liverwort Lepidolaena taylorii is the known ent-8,9-seco-7-bydroxy-kaura-8(14),16-dien-9,15-dione 1. The relative stereochemistry of a new cytotoxic compound, 16,17-dihydro derivative 2, was supported by molecular modelling. Another new cytotoxic compound was identified as 8,14-epoxide 3 by preparation from 1. These compounds showed differential cytotoxic activity against human tumor cell lines. Copyright © 1996 Elsevier Science Ltd

We are searching for bioactive natural products from New Zealand's native plants, focusing on liverworts because of their rich array of secondary metabolites. Crude extracts of the liverwort *Lepidolaena taylorii* (Gott.) Trev. (family Lepidolaenaceae) showed strong cytotoxic activity in P388 leukemia assays. There are no published reports of *Lepidolaena* chemistry. Bioactivity-directed fractionation led to three cytotoxic 8,9-secokauranes 1 - 3. This is the first report of 8,9-secokauranes from any source other than the higher plant genus *Rabdosia* (family Lamiaceae). The two most active compounds, rabdoumbrosanin 1 and epoxide 3, showed differential cytotoxicity in the human disease oriented *in vitro* assays of the US National Cancer Institute (NCI).

An extract of *L. taylorii* (4.05 g, P388 IC₅₀ 1.3 μ g/ml)⁷ subjected to reverse phase (C18) flash chromatography gave cytotoxic activity (IC₅₀'s < 0.5 μ g/ml) spread across fractions eluted with 1:1, 1:3 and 1:9 H₂O:MeCN. Silica TLC showed that these fractions were mostly complex mixtures. One of the most cytotoxic fractions (96 mg, 1:3 H₂O:MeCN, IC₅₀ 0.06 μ g/ml), which contained one major compound by ¹H NMR, was fractionated by silica column chromatography and C18 HPLC.⁸ The major compound 1 was finally separated from a minor (10%) compound 2 by changing the C18 HPLC mobile phase from H₂O:MeCN to H₂O:MeOH.⁹

The high resolution MS of compound 1 was consistent with the molecular formula $C_{20}H_{28}O_3$.¹⁰ The IR, ¹H and ¹³C NMR spectra suggested the presence of -OH, two ketone C=O, a trisubstituted C=C bond, a 1,1-disubstituted C=CH₂, and three methyl singlets.¹⁰ A search of the Chemical Abstracts Registry file retrieved only one compound which matched all these structural requirements. The spectral data, especially ¹³C NMR shifts,

reported for rabdoumbrosanin 1 matched our cytotoxic compound from *L. taylorii*, 5,10 and the optical rotations showed the same absolute stereochemistry. Rabdoumbrosanin 1 is an *ent-8*,9-secokaurane diterpenoid which has only been reported once before, with no mention of biological activity, from the higher plant *Rabdosia umbrosa* (Maxim.) Hara. 5

The MS of minor compound 2 was similar to the MS of 1, but with the higher m/z ions all +2 Daltons and a molecular ion consistent with the formula $C_{20}H_{30}O_3$.¹¹ The ¹H NMR spectrum of 2 showed a methyl doublet, and lacked the signals for a 1,1-disubstituted C=CH₂ group,¹¹ suggesting that this compound was a 16,17-dihydro derivative 2 of rabdoumbrosanin 1. No 16,17-dihydro-8,9-secokauranes have been reported previously, although 16,17-dihydrokaurane derivatives have been found in *Rabdosia* species.⁴ The ¹³C NMR spectrum of 2¹¹ showed signals within 1 ppm of the signals assigned to C-1 to C-11 and C-18 to C-20 of 1.⁵ Therefore 2 was assigned the same relative stereochemistry as 1 at C-5, C-7, C-10 and C-13. The stereochemistry at C-16 of 2 was assigned by

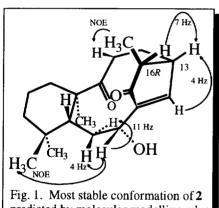


Fig. 1. Most stable conformation of 2 predicted by molecular modelling, plus selected NMR interactions.

comparing proton NOE interactions and coupling data with the results of molecular modelling. Conformational searching and molecular mechanics minimisations¹² were used to predict the most stable conformations of the C-16 R and S configurations of 2. The NMR data, particularly the H-16 to H-13 coupling of 7 Hz, best matched the C-16 R configuration of 2 (Fig. 1).

In order to obtain more of compound 1 for biological testing (see below), a second cytotoxic fraction from C18 flash chromatography (220 mg, 1:3 $\rm H_2O$:MeCN, IC₅₀ 0.04 $\rm \mu g/ml$) was subjected to preparative C18 HPLC.¹³ In addition to 1 and 2, compound 3 was obtained pure. High resolution MS¹⁴ was consistent with the molecular formula $\rm C_{20}H_{28}O_4$, with one more oxygen atom than 1. The ¹H NMR spectrum of 3 was similar to

that of 1, but lacked the low-field H-14 signal and showed an additional singlet at 3.71 ppm. This suggested that 3 was the previously unreported 8,14-epoxide of 1. This was proved by epoxidation of 1, which occurred selectively at the trisubstituted 8,14 bond, 12 to give 3.15 Three 8,14-epoxy-8,9-secokauranes have been reported, and showed H and 13C NMR signals similar to 3.34 Molecular modelling of 8,9-secokauranes (Fig. 1) suggested

epoxidation was only possible on the "outside" face of the cyclopentenone to give the 14R stereochemistry shown for 3.

All three 8,9-secokauranes from L. taylorii showed strong cytotoxic activity against P388 leukemia cells: 1 had an IC₅₀ of 0.06 µg/ml (SD \pm 0.02, n = 2), equivalent to 0.2 µM; 2 had an IC₅₀ of 0.8 µg/ml (SD \pm 0.2, n = 2); and 3 had an IC₅₀ of 0.27 µg/ml (SD \pm 0.06, n = 4). The *in vitro* cytotoxicity and *in vivo* antitumor activity reported for other 8,9-secokauranes has been ascribed to Michael addition of bio-nucleophiles to the α , β -unsaturated ketone group.³ If this is the mode of action, it seems that either the exocyclic or the endocyclic double bond can undergo Michael addition, since both 16,17-dihydro derivative 2 and 8,14-epoxide 3 are cytotoxic. Compound 1 (containing 10% of 2) and compound 3 showed differential cytotoxicity in *in vitro* human tumor assays.⁶ Growth of five leukemia cell lines was inhibited (mean IC₅₀ of 0.4 µM for 1) but the cells were not killed (mean LC₅₀ > 50 µM). By contrast, other tumor cell lines were killed. For example, the mean LC₅₀ of 1 against seven colon cancer cell lines was 6 µM. Compounds 1 and 3 are now being evaluated for *in vivo* antitumor activity. We are continuing investigations of the other cytotoxic compounds in L. taylorii.

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- Collected from Bruce Bay, West Coast, South Island, New Zealand in June 1995 (University of Otago Herbarium number OTA 6803). Dried liverwort (100 g) was extracted with EtOH then CHCl₃. For the P388 assay method, see Lorimer et al..¹
- Flash chromatography on silica with 3:1 cyclohexane:EtOAc; followed by C18 HPLC (Merck Lichrospher 100 RP-18 10 μm, 250 x 10 mm, mobile phase 5 ml/min 1:3 H₂O:MeCN, detection at 206 nm): peak retention time (RT) 4.7 min (1 plus 2, 12 mg).
- 9. C18 HPLC as above except with 3:7 H₂O:MeOH: peak RTs 10.8 min (1) and 12.4 min (2).

- 10. *Rabdoumbrosanin (Registry No. 123941-77-5) I.* colourless oil; silica TLC R_F 0.3 (2:1 cyclohexane:EtOAc, UV plus yellow with anisaldehyde dip); $[\alpha]^{21}_{D}$ -66° (c 0.5, MeOH); UV (MeOH) λ max ($\log \epsilon$) 245 (3.71) nm; IR (film) v max 3442 (br, OH), 2948 (CH), 1694 (both C=O), 1646 (C=C) cm⁻¹; 1 H NMR (CDCl₃) δ 7.24 (1 H, br d, 2 Hz, H-14), 6.12 (1 H, br s, H-17), 5.43 (1 H, s, H-17), 4.68 (1 H, dd, 12, 5 Hz, H-7), 3.59 (1 H, br m, H-13), 2.58 (1 H, td, 12, 4 Hz, H-12), 2.3 (1 H, m, H-11), 1.9-1.1 (m), 1.00 (3 H, s), 0.94 (3 H, s), 0.92 (3 H, s), 0.88 (1 H, dd, 6, 2 Hz, H-5); 13 C NMR (CDCl₃) δ 215.3 (s, C-9), 195.2 (s, C-15), 159.7 (d, C-14), 148.5 (s, C-16), 146.0 (s, C-8), 116.9 (t, C-17), 64.4 (d, C-7), 53.9 (s, C-10, 43.5 (d, C-5), 42.4 (d, C-13), 41.4 (t, C-3), 36.7 (t, C-6), 34.7 (s, C-4), 34.1 (t, C-1), 33.7 (q, C-18), 30.8 (t, C-11), 25.9 (t, C-12), 22.4 (q, C-19), 18.1 (t, C-2), 16.6 (q, C-20); EIMS (70 eV) m/z 316.2042 (M^+ , 3%, $C_{20}H_{28}O_3$ req 316.2038), 301 (2), 298 (9), 283 (4), 242 (6), 192 (48), 179 (100), 151 (18), 138 (31), 123 (70).
- 11. 16.17-Dihydro-rabdoumbrosanin 2.- colourless oil; silica TLC R_F 0.2 (2:1 cyclohexane:EtOAc, purple spot with anisaldehyde dip); $[\alpha]_D^{21}$ -11° (c 0.15, CHCl₃); UV (MeOH) λ max (log ϵ) 213 (4.58) nm; IR (film) ν max 3436 (br, OH), 2942 (CH), 1697 (both C=O) cm⁻¹; 1 H NMR (CDCl₃) δ 7.34 (1 H, br d, 3 Hz, H-14), 4.58 (1 H, dd, 12, 5 Hz, H-7), 3.20 (1 H, br m, H-13), 2.50 (1 H, quintet, 7 Hz, H-16), 2.42 (1 H, m, H-12), 2.02 (1 H, br dd, 18, 13 Hz, H-11), 1.9-1.2 (m), 1.14 (3 H, d, 7 Hz, H-17), 1.04 (3 H, s, H-18), 0.95 (3 H, s, H-20), 0.92 (3 H, s, H-19); 13 C NMR (CDCl₃) δ 214.4 (s, C-9), 210.0 (s, C-15), 162.7 (d, C-14), 145.3 (s, C-8), 64.3 (d, C-7), 53.8 (s, C-10), 44.7 (d, C-16), 42.7 (d, C-5), 42.3 (d, C-13), 41.3 (t, C-3), 36.9 (t, C-6), 34.5 (s, C-4), 34.3 (t, C-1), 33.4 (q, C-18), 31.4 (t, C-11), 22.3 (q, C-19), 21.2 (t, C-12), 18.1 (t, C-2), 16.4 (q, C-20), 10.0 (q, C-17); EIMS (70 eV) m/z 318.2192 [M]⁺ (3%,C₂₀H₃₀O₃ req 318.2195), 300 (6), 244 (12), 194 (24), 181 (100), 138 (55), 123 (99).
- 12. For modelling methods, see Hinkley, S. F.; Perry, N. B.; Weavers, R. T. Phytochemistry 1994, 35, 1489-1494.
- 13. C18 HPLC as above except with 2:3 H₂O:MeCN: peak RTs 7.4 min (1 plus 2, 18 mg) and 9.7 min (3, 26 mg).
- 14. 8,14-Epoxy-rabdoumbrosanin 3.- colourless oil; silica TLC R_F 0.5 (2:1 cyclohexane:EtOAc, UV spot); $[\alpha]^{25}_D$ -53° (c 0.46, MeOH); UV (MeOH) λ max ($\log \epsilon$) 235 (3.57) nm; IR (film) ν max 3490 (br, OH), 2951 (CH), 1726 (C-15=O), 1694 (C-9=O), 1643 (C=C) cm⁻¹; 1 H NMR (CDCl₃) δ 6.29 (1 H, br s, H-17), 5.53 (1 H, d, 1 Hz, H-17), 4.61 (1 H, dd, 12, 5 Hz, H-7), 3.71 (1 H, s, H-14), 3.22 (1 H, br m, H-13), 2.65 (2 H, m), 2.0-1.1 (m), 1.06 (3 H, s), 0.99 (3 H, s), 0.94 (3 H, s); 13 C NMR δ 214.5 (s, C-9), 196.9 (s, C-15), 145.0 (s, C-16), 123.0 (t, C-17), 64.6 (s, C-8), 61.9 (d, C-14), 60.6 (d, C-7), 53.7 (s, C-10), 42.0 (d, C-5), 41.5 (t, C-3), 40.0 (d, C-13), 34.9 (s, C-4), 33.9 (t, C-6), 33.4 (q, C-18), 33.2 (t, C-1), 31.4 (t, C-11), 25.2 (t, C-12), 22.0 (q, C-19), 18.1 (t, C-2), 16.4 (q, C-20); EIMS (70 eV) m/z 332.1987 (M⁺, 1%, $C_{20}H_{28}O_4$ req 332.1988), 314 (5), 299 (3), 258 (5), 209 (17), 195 (8), 192 (8), 179 (20), 178 (14), 165 (10), 150 (18), 149 (11), 138 (11), 137 (20), 123 (100).
- 15. m-Chloroperoxybenzoic acid (2.7 mg) was added to 1 (5 mg) dissolved in CH₂Cl₂ (1 ml), and stirred with aq NaHCO₃ (0.5 M, 0.3 ml). After 2 days, silica TLC and ¹H NMR spectroscopy showed conversion to 3.